

**SOP:** Propagation of Mouse MEL-GATA-1-ER cells  
**Date modified:** 01/12/2011  
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### **Ordering Information**

Mouse MEL-GATA-1-ER cells (mG/ER) were received from Dr. Arthur Skoultchi, Albert Einstein College of Medicine of Yeshiva University, Bronx, New York.

### **Notes:**

This is a mouse suspension cell line derived from MEL cells by stable transfection with a GATA-1-ER fusion protein construct as described by Choe et al., 2003 (Cancer Res 63, 6363–6369, 2003). These cells can be terminally differentiated into mature erythroid cells with  $\beta$ -estradiol treatment.

### **Materials List**

1. DMEM, 1X, Dulbecco's Modification of Eagle's Medium, with 4.5g/L glucose, L-glutamine, sodium pyruvate (Cellgro, Cat# 10-013-CV)
2. Characterized Fetal Bovine Serum (HyClone, Cat# SH-30071-03)
3. Penicillin-Streptomycin Solution, 200X (Cellgro, Cat# 30-001-CI)
4. Phosphate Buffered Saline (1X PBS) (Cellgro, Cat# 21-040-CM)
5. Puromycin dihydrochloride (Sigma-Aldrich, Cat# P8833)
6.  $\beta$ -Estradiol (Sigma-Aldrich, Cat# E2257)
7. T75, T225 tissue culture flasks
8. Corning conical centrifuge tubes (15mL and 50mL)
9. Graduated serological pipets (1, 5, 10, 25, 50mL)
10. Freezing Medium (growth medium containing 10% DMSO)
11. DMSO, Hybri-Max (Sigma-Aldrich, Cat# D2650)
12. Cryo Vials (Nunc, Cat# 368632)
13. Cryo 1°C Freezing Container (Nalgene, Cat# 5100-0001)
14. Eppendorf Centrifuge 5810R
15. Revco UltimaII -80°C Freezer
16. Thermolyne Locator 4 Liquid Nitrogen Freezer
17. Hemocytometer
18. Micropipet w/ P20 tips
19. Microscope

### **Growth Medium for Mouse G/ER Cells**

DMEM, 1X, with 4.5g/L glucose, L-glutamine, sodium pyruvate  
10% Characterized FBS  
Pen-Strep (1X)  
Puromycin dihydrochloride (5 $\mu$ g/mL)

### **Procedure**

#### **A. Receipt of Frozen Cells and Starting Cell Cultures**

1. Immediately place frozen cells in liquid nitrogen storage until ready to culture.
2. When ready to start cell culture, quickly thaw ampoule in 37°C water bath until ice crystals disappear.
3. Swab outside of the ampoule with 70% ethanol and then dispense contents of ampoule into a 15mL Corning conical centrifuge tube.
4. Add 10mL cold growth medium, drop wise, into the centrifuge tube containing cells.

5. Pellet cells gently at 200 x g 4°C 5 minutes and remove DMSO-containing supernatant.
6. Resuspend pellet at  $2 \times 10^5$  cells/mL with pre-warmed growth medium and grow in a 37°C, 10% CO<sub>2</sub> humidified incubator. **Concentration of cells should never exceed  $1 \times 10^6$  cells/mL.**

#### **B. Sub-culture and Maintenance**

1. Take cell counts with a hemocytometer every 24 hours to maintain the culture at a cell density between  $2 \times 10^5$  cells/mL and  $1 \times 10^6$  cells/mL. The cells have a doubling time of 12 hours and the concentration of cells should not exceed  $1 \times 10^6$  cells/mL.
2. Add fresh warm medium when appropriate to maintain cell density and expand the culture to the desired number of cells.
3. Record each subculture event as a passage.

#### **C. Generation of Seed Stocks**

1. At an early stage of expansion and with sufficient number of cells to continue maintenance, a small portion of the cells should be set aside as a seed stock, if needed.
2. Amount of cells for the seed stock should be placed in a conical centrifuge tube and centrifuged at 500 x g (4°C) for 5 minutes.
3. Aspirate supernatant and resuspend the cell pellet in 1X PBS to wash. Centrifuge again under same conditions.
4. Resuspend the cell pellet in freezing medium (growth medium containing 10% DMSO) at a concentration yielding 2 million cells per 1mL aliquot.
5. Dispense 1mL cell suspension per cryovial. Place cryovials in a Nalgene Cryo 1°C freezing container and store overnight at -80°C.
6. Cryovials are transferred the next day to liquid nitrogen freezer for long-term storage.

#### **D. Harvest**

1. Passage cells until the desired number of cells for experimentation is reached in a logarithmic growth phase.
2. Pellet cells and rinse with 1X PBS as in “Generation of Seed Stocks” section.
3. Examine viability using Trypan blue staining (SOP TP-7).

#### **E. Differentiation**

1. The cells can be differentiated to red blood cells by adding  $\beta$ -estradiol to the growth medium to  $10^{-7}$ M final concentration (freshly thawed  $10^{-4}$ M stock from -20°C storage, made according to Sigma-Aldrich specifications for solubilization and storage). The cells are committed to differentiation within 48 hours; within 120 hours, 95% of cells are differentiated.